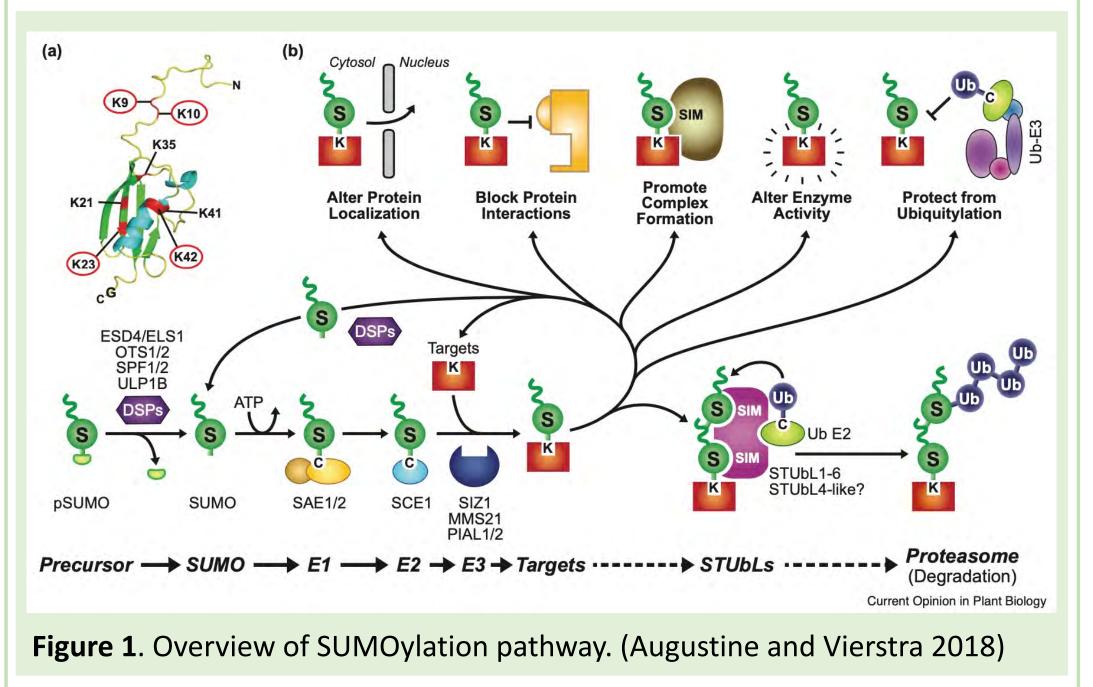
CRISPR/Cas9 Mediated Mutagenesis of Moss SUMO System & Development of a Decahistidine(10His)-tagged SUMO Purification Line

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Introduction & Background

SUMOylation

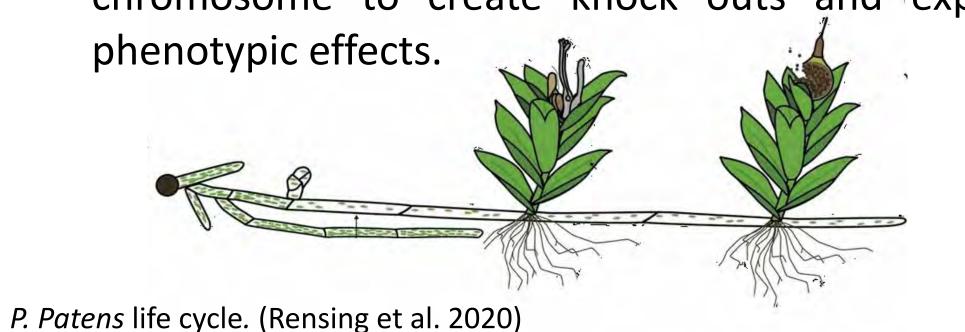
- Small Ubiquitin-related Modifier (SUMO) is a small protein that acts as post translational modifier affecting protein activity, localization, and stability.
- SUMOylation (the process of SUMO attachment) occurs within minutes of expose to environmental stressors, providing resilience ensuring survival under suboptimal conditions.
 - Four main steps of the SUMO cycle include: Activation (E1 - SAE1/2) , Conjugation (E2 -SCE1), Ligation (E3 - SIZI1), and DeSUMOylation by DPS's



CRISPR modifies target areas of the genome and can knock out genes of interest in the SUMO pathway to better characterize the system.

The SUMO system in *Physcomitrium patens*

- Highly amenable to genetic and bioanalysis.
- Haploid cells require modification of only one chromosome to create knock outs and express phenotypic effects.



Goals

- Genetically interrogate the SUMO system in Physcomitrium patens and identify the proteins modified by SUMO.
- > Isolation and purification of a 10His-SUMO1a line for identification of SUMOylated proteins.

Selected References

1. Augustine, Robert C., and Richard D. Vierstra. "SUMOylation: Re-Wiring the Plant Nucleus during Stress and Development." Current Opinion in Plant Biology, vol. 45, Oct. 2018, pp. 143-54. 2. Hendriks, Ivo A, and Alfred C. O. Vertegaal. "A High-Yield Double-Purification Proteomics Strategy for

the Identification of SUMO Sites." Nature Protocols, vol. 11, no. 9, Sept. 2016, pp. 1630-49. 3. Hendriks, Ivo A., and Alfred C. O. Vertegaal. "Label-Free Identification and Quantification of SUMO Target Proteins." SUMO, edited by Manuel S. Rodriguez, vol. 1475, Springer New York, 2016, pp. 171–93. 4. Peroutka III, Raymond J., et al. "SUMO Fusion Technology for Enhanced Protein Expression and Purification in Prokaryotes and Eukaryotes." Heterologous Gene Expression in E.Coli, edited by Thomas C. Evans, and Ming-Qun Xu, vol. 705, Humana Press, 2011, pp. 15-30.

5. Rensing, Stefan A., et al. "The Moss Physcomitrium (Physcomitrella) Patens: A Model Organism for Non-Seed Plants." The Plant Cell, vol. 32, no. 5, May 2020, pp. 1361–76. 6. Rodriguez, Manuel S., editor. SUMO: Methods and Protocols. Springer New York, 2016.



QR Code of Audio File

Acknowledgements

Thank you to the Vassar College, the Biological Science Department, and URSI program for sponsoring and providing the opportunity to complete this project. A special thank you to Professor Augustine for the guidance, experience, and everything I learned along the way. Thank you to David Lewis for all your support and help throughout the project.

Decahistine (10His)-tag Development Methods Co-transformation of SUMO1a CRISPR/Cas9 with 10His replacement construct to induce Homology Directed repair growing on antibiotic-containing Electrophoresis Sanger Sequencing to confirm insertion of 10His Grind and propogate selected moss 2 3 4 5 6 7 8 9 Selection of Moss with potential 10His-tag insertion Heat Stress at 38C Protein Extraction Purification of SUMO via Nickel Column Western Blot of 10His and WT

Results 7-30 Decahistadine Wild Type CE FT W1 EL1 EL2 EL2 EL1 W1 FT CE @SUMO

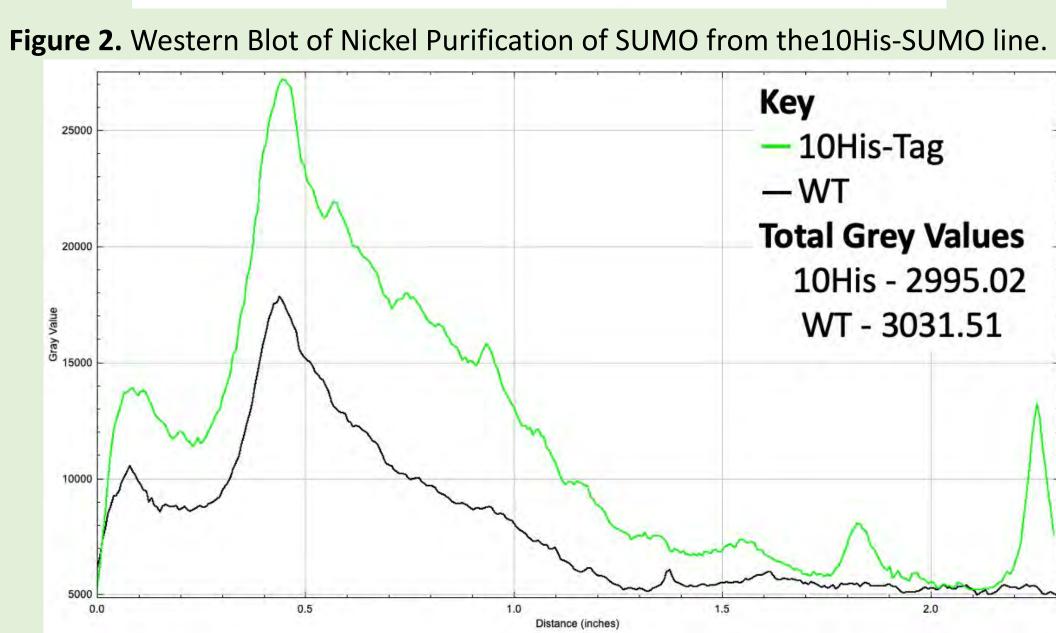


Figure 3. Quantitative comparison of grey values from lanes EL1 of WT vs. 10His tagged moss from the western blot (Figure 2).

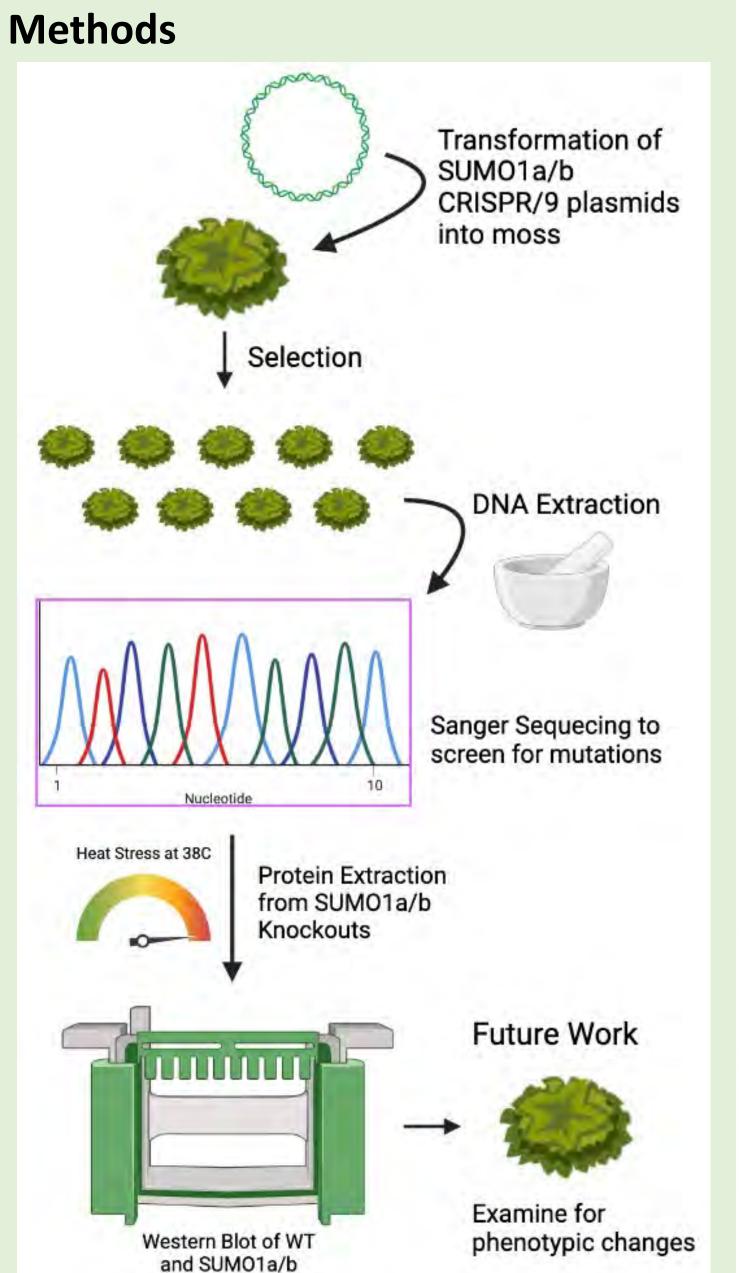
Conclusions

The 10His-tag enables purification of free SUMO and SUMO1a conjugates thereby enabling their proteomic identification.

Future Work

 Creation of a line where both SUMO1a and SUMO1b are 10His tagged.

CRISPR/Cas9 Mediated Mutagenesis



Knockouts



Figure 4. Amino acid sequence alignment of predicted sequences of wild type and selected SUMO1b mutants. DiGlycine (highlighted by the red box) residues are required for SUMOylation.

SUMO1a Mutants SUMO1b Mutants 130-100-70 — 40 35 -20 15-

Figure 5. Western Blot of SUMO protein extracted from moss with SUMO1a and SUMO1b genes knockouts.

Conclusions

SUMO1a and SUMO1b genes encode for distinct variants of the SUMO protein.

Future Work

- Analysis of SUMO1a/b knockouts on the phenotype.
- Further exploration of SUMO variants.